Denaturing Protocol
1. Titrate the sample with HCl to bring the pH to 2
2. Add SDS (final concentration should be 0.1%) and $\beta$-mercaptoethanol (final concentration 100mM)
   - Check the fluorescence on the blue box
3. Separate the strand from the LOO-GFP with the centricon (4000 rpm for 15 minutes)
4. Add SDS buffer 1 to the centricon and spin at 4000 rpm for 15 minutes
5. Titrate the solution to a pH of 8 with SDS buffer 2 (keep the SDS concentration at 0.1% and the $\beta$-mercaptoethanol at 100mM)
6. Wash in centricon with 0.1% CHAPS and 10mM $\beta$-mercaptoethanol
   - Check the fluorescence on the blue box
   - Run fraction on FPLC
7. Wash with TN buffer
   - Run fraction on FPLC
   - Check the fluorescence on the blue box

Future Work: Characterize samples before denaturation, after denaturation, after refolding, after the addition of the peptide

SDS buffer 1-0.1% SDS, 10mM $\beta$-mercaptoethanol, TN buffer, pH2
SDS buffer 2-0.1% SDS, 10mM $\beta$-mercaptoethanol, TN buffer, pH8
CHAPS buffer – 0.1% CHAPS, 10 mM $\beta$-mercaptoethanol, TN buffer pH 8
TN buffer – 100 mM NaCl, 50 mM Tris